Connexins and pannexins

Overview: Gap junctions are essential for many physiological processes including cardiac and smooth muscle contraction, regulation of neuronal excitability and epithelial electrolyte transport (see Evans and Martin, 2002; Bruzzone et al., 2003; Connors and Long, 2004). Gap junction channels allow the passive diffusion of molecules of up to 1000 Da that can include nutrients, metabolites and second messengers (such as IP₃) as well as cations and anions. Twenty-one connexin genes (Cx23, Cx25, Cx26, Cx30, Cx30.2, Cx30.3, Cx31, Cx31.1, Cx31.9, Cx32, Cx36, Cx37, Cx40, Cx40.1, Cx43, Cx45, Cx46, Cx47, Cx50, Cx59 and Cx62) and three pannexin genes (Px1, Px2 and Px3; which are structurally related to the invertebrate innexin genes) code for gap junction proteins in humans. Each connexin gap junction comprises two hemichannels or 'connexons' that are themselves formed from six connexin molecules. The various connexins have been observed to combine into both homomeric and heteromeric combinations, each of which may exhibit different functional properties. It is also suggested that individual hemichannels formed by a number of different connexins might be functional in at least some cells (see Herve et al., 2007). Connexins have a common topology, with four α-helical transmembrane domains, two extracellular loops, a cytoplasmic loop and N- and C-termini located on the cytoplasmic membrane face. In mice, the most abundant connexins in electrical synapses in the brain seem to be Cx36, Cx45 and Cx57 (Sohl et al., 2005). Mutations in connexin genes are associated with the occurrence of a number of pathologies, such as peripheral neuropathies, cardiovascular diseases and hereditary deafness. The pannexin genes Px1 and Px2 are widely expressed in the mammalian brain (Vogt et al., 2005). Like the connexins, at least some of the pannexins can form hemichannels (Bruzzone et al., 2003; Pelegrin and Surprenant, 2007).

Connexins

Nomenclature

Cx23, Cx25, Cx26, Cx30, Cx30.2, Cx30.3, Cx31, Cx31.1, Cx31.9, Cx32, Cx36, Cx37, Cx40, Cx40.1, Cx43, Cx45, Cx46, Cx47, Cx50, Cx59, Cx62

Ensembl ID

ENSG00000159248 (Cx36)*

Inhibitors

Carbenoxolone Flufenamic acid Octanol

Raising external calcium

Pannexins

Px1, Px2, Px3

ENSG00000110218 (Px1) ENSG00000073150 (Px2) ENSG00000154143 (Px3)

Carbenoxolone

Little block by flufenamic acid Unaffected by raising external calcium

Connexins are most commonly named according to their molecular weights, so, for example, Cx23 is the connexin protein of 23 kDa. This can cause confusion when comparing between species - for example the mouse connexin Cx57 is orthologous to the human connexin Cx62. No natural toxin or specific inhibitor of junctional channels has been identified; however, two compounds often used experimentally to block connexins are carbenoxolone and flufenamic acid (Salameh and Dhein, 2005). At least some pannexin hemichannels are more sensitive to carbenxolone than connexins but much less sensitive to flufenamic acid (Bruzzone et al., 2005). It has been suggested that 2-aminoethoxydiphenyl borate (2-APB) may be a more effective blocker of some connexin channel subtypes (Cx26, Cx30, Cx36, Cx40, Cx45 and Cx50) compared with others (Cx32, Cx43 and Cx46, Bai et al., 2006).

*Due to space constraints, the Ensembl ID for only Cx36 is given. Ensembl information for the other connexins can be found from links therein.

Further Reading

Bennett MV, Zukin RS (2004) Electrical coupling and neuronal synchronization in the mammalian brain. Neuron 41: 495-511.

Connors BW, Long MA (2004). Electrical synapses in the mammalian brain. Ann Rev Neurosci 27: 393-418.

Cruciani V, Mikalsen SO (2006). The vertebrate connexin family. Cell Mol Life Sci 63: 1125-1140.

Evans WH, Martin PEM (2002). Gap junctions: structure and function. Mol Memb Biol 19: 121-136.

Evans WH, De Vuyst E, Leybaert L (2006). The gap junction cellular internet: connexin hemichannels enter the signalling limelight. Biochem J

Herve JC, Sarrouilhe D (2005). Connexin-made channels as pharmacological targets. Curr Pharm Des 11: 1941–1958.

Herve JC, Phelan P, Bruzzone R, White TW (2005). Connexins, innexins and pannexins: Bridging the communication gap. Biochim Biophys Acta

Homuzdi SG, Filippov MA, Mitropoulou G, Monyer H, Bruzzone R (2004). Electrical synapses: a dynamic signaling system that shapes the activity of neuronal networks. Biochim Biophys Acta 1662: 113–137.

Kumar NM, Gilula NB (1996). The gap junction communication channel. Cell 84: 381-388.

Mese G, Richard G, White TW (2007). Gap junctions: basic structure and function. J Invest Dermatol 127: 2516-2524.

Salameh A, Dhein S (2005). Pharmacology of gap junctions: New pharmacological targets for treatment of arrhythmia, seizure and cancer? Biochim Biophys Acta 1719: 36-58.

Shestopalov VI, Panchin Y (2008). Pannexins and gap junction protein diversity. Cell Mol Life Sci 65: 376–394.

Sohl G, Maxeiner S, Willecke K (2005) Expression and functions of neuronal gap junctions. Nature Rev Neurosci 6: 191-200.

Spray DC, Dermietzel R (1996). Neuroscience Intelligence Unit: Gap Junctions in the Nervous System. Springer: New York.

Yen MR, Saier MH (2007). Gap junctional proteins of animals: the innexin/pannexin superfamily. Prog Biophys Mol Biol 94: 5-14.

References

Bai D et al. (2006). J Pharmacol Exp Ther 319: 1452-1458. Bruzzone R et al. (2003). Proc Natl Acad Sci USA 100: 13644-13649. Bruzzone R et al. (2005). J Neurochem 92: 1033-1043.

Herve JC et al. (2007). Prog Biophys Mol Biol 94: 29-65. Pelegrin P, Surprenant A (2007). J Biol Chem 282: 2386–2394. Vogt A et al. (2005). Brain Res Mol Brain Res 141: 113-120.